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Legume-based mixed intercropping systems may lower agricultural born N₂O emissions

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Abstract

Background: The area used for bioenergy crops (annual row crops (e.g., wheat, maize), herbaceous perennial grasses, and short-rotation woody crops (e.g., poplar)) is increasing because the substitution of fossil fuels by bioenergy is promoted as an option to reduce greenhouse gas (GHG) emissions. However, biomass used for bioenergy production is not per se environmentally benign, since bioenergy crop production is associated with negative side effects such as GHG emissions from soil (dominated by N₂O). N₂O emissions vary greatly in space and time; thus, direct comparison of soil N₂O fluxes from various agro-ecosystems is certainly crucial for the assessment of the GHG reduction potential from energy crops.

Methods: Therefore, our study aimed to evaluate the two different agro-ecosystems (cropland and agro-forestry) cultivated in central Germany for their environmental impact. In a 1-year field experiment, we compared N₂O fluxes from cropland (non-fertilized wheat, N-fertilized wheat, non-fertilized faba bean, and wheat mixed intercropping with faba bean) and agro-forestry (non-fertilized poplar, N-fertilized poplar, non-fertilized Robinia, and poplar mixed intercropping with Robinia) as a randomized split-block design.

Results: Rainfall at the field site was slightly over average during the period from 1 April to 1 July in 2014 (201 mm rain) and considerably below average during the same period in 2015 (100 mm rain). Cumulative mean N₂O fluxes were up to five fold higher in agro-forestry than in arable crop treatments during 2014 growing period. We hypothesized that the difference in N₂O emissions when comparing arable land and agro-forestry was mainly due to the limited water and nutrient uptake of plantations during the first year. Among the arable crops (wheat, N-fertilized wheat, wheat mixed intercropped with bean, and bean), seasonal and annual N₂O emissions were highest in soils when faba bean was grown as a mono-crop. On the other hand, cumulative mean N₂O fluxes were 31 % lower ($p < 0.05$) when faba bean mixed with wheat than in soils planted with N-fertilized wheat.

Conclusions: The latter clearly suggests that using legume crops as intercrop or mixed crop in wheat may significantly mitigate fertilizer-derived N₂O fluxes and may be an effective proxy for increasing GHG emission savings for energy crops.

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Background

The observed increase in global average temperatures over the last decades is very likely due to the observed increase in anthropogenic greenhouse gas concentrations in the atmosphere. Nitrous oxide (N_2O) is a potent greenhouse gas as it absorbs long-wave radiation and contributes to the reduction of the ozone layer in the stratosphere [1]. Data from the ice-core analysis show that for thousands of years, mean atmospheric N_2O concentrations were close to 270 ppbv; however, the latter increased about 20 % in recent years [2, 3]. The Intergovernmental Panel on Climate Change (IPCC) Third Assessment Report identified microbial production of N_2O in expanding and fertilized agricultural lands as a primary driver of this increase [4]. However, large uncertainties remain on the estimates of N_2O fluxes from the biosphere [2, 5] due to complex interactions between the related processes and controls on production, consumption, and transport through the soil and on the release into the atmosphere [6, 7].

High-yielding agricultural systems have specifically high nitrogen (N) demand which cannot be supplied by soil N reserves. Therefore, additional N input is needed; however, in most agricultural land-use systems, the application of organic and inorganic nitrogen fertilizers triggers the emissions of anthropogenic N_2O emissions. The present IPCC default factor for direct N_2O emissions arising from the nitrogenous fertilizer application to managed soils is 1 %. However, soil N_2O emissions vary significantly depending on soil type, plant species, climate, crop rotation, tillage method, and fertilizer application rates [4]. At high N_2O -emitting sites, most N_2O release is characterized by short peak emissions (up to 90 % of the annual emissions) connected mainly to (i) precipitation events and change in soil moisture [8–10], (ii) N fertilization [11–13], (iii) freeze-thaw cycle [14], and (iv) soil tillage [15]. Such peak N_2O emissions from soils are highly variable in space and time; thus, measuring and quantifying variance in N_2O emissions are rather difficult, and there are only a few field experiments available allowing long-term comparison of various crops and other factors [16]. It is absolutely crucial to simulate such N_2O peak events and to identify the driving factors in order to be able to develop mitigation options.

The main microbial reactions involved in the production of N_2O are nitrification (oxidation of NH_4^+ to NO_3^-) and denitrification (reduction of NO_3^- , via NO , N_2O , to N_2). There is now growing evidence that denitrification (bacterial or fungal) is the dominating process responsible for N_2O losses from agricultural soils [8, 10, 17]. Therefore, in many agricultural field studies, N_2O emission events are found in periods when high mineral nitrogen concentrations in soils coincide with high soil moisture (increasing the rate of denitrification) [7, 18].

The relevance of soil moisture can also be seen in field studies with similar climatic, soil, and substrate conditions, where N_2O emissions nevertheless show significant inter-annual variability caused by the differences in soil water content [19]. These differences can usually be deduced from the weather records as they are related to the differences in the input of water through rainfall [18]. But the soil-water balance is also strongly influenced by the offtake of water through evapotranspiration. Crop plants play an important role, as they may transpire 500 to 600 mm of water per growth cycle which is 30 to 90 % of the precipitation input in that period. In this respect, crops differ substantially, not only in the total amount of water that is transpired but also in the growth and transpiration pattern in the course of the year [19, 20].

Biofuels are often called “ CO_2 neutral” in the sense that CO_2 which is emitted in the course of their combustion has previously been fixed from the atmosphere via photosynthesis during plant growth. Many industrialized countries have established ambitious policy targets and often offer financial incentives to stimulate the production or use of bioenergy. The main reasons for the promotion of biofuel production are that it is made from renewable resources (organic manures, plant materials, food waste), that it is expected to have no or even positive effect on the atmospheric greenhouse gas balance, and that it may reduce the dependency on fossil fuel [21]. However, biomass used for bioenergy generation is not per se environmentally benign, since its production is inevitably associated with negative side effects such as GHG emissions or N leaching. Soil N_2O emissions are likely to be the dominating greenhouse gas emissions associated with bioenergy crop production [22]. It has been reported that the production and use of biofuels compared with the use of conventional fossil fuels may lead to a reduction or even increase in the total greenhouse gas emissions (72 to 107 %), depending on the type of bioenergy crop used and combustion technology chosen [23]. Here, the authors showed that N_2O would typically make up 10 to 80 % of the total greenhouse gas emissions in the biofuel production chain.

Intercropping, defined as any system of multiple cropping within the same space can be used as an alternate bioenergy cropping system [24]. The intercropping of cereals with legumes is particularly common, and introducing N_2 -fixing legumes into cereal-based crop rotations may reduce synthetic mineral N-fertilizer use and thought to mitigate N_2O fluxes. A reduction of N_2O in tree-based intercropping systems has been reported [25]. In contrast, in a review study, Rochette and Janzen [26] concluded that legumes can produce substantial N_2O emissions. The cultivation of N_2 -fixing legume species (e.g., faba bean (*Vicia faba* L.) as arable crop or Robinia

(*Robinia pseudoacacia*) as woody plant) could stimulate N_2O emissions simply by increasing N input to soils, thus providing additional substrate for nitrification and denitrification [27]. Authors reported that faba bean could release 13 % of their fixed N as rhizodeposition [28]. Furthermore, denitrification by N-fixing bacteria can be another source of N_2O in legume domains [29].

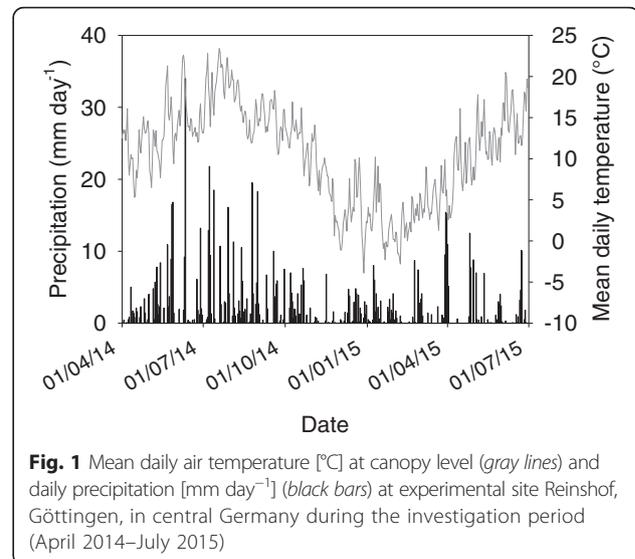
As discussed above, N_2O emissions vary greatly in space and time; thus, the direct comparison of soil N_2O fluxes from various agro-ecosystems is certainly necessary and only very few studies are available on this subject. Therefore, our study aimed to evaluate two different agro-ecosystems (cropland and agro-forestry) cultivated in central Germany for their environmental impact. In a 1-year field experiment, we compared N_2O fluxes from cropland (non-fertilized wheat, N-fertilized wheat, non-fertilized faba bean, and wheat mixed intercropping with faba bean) and agro-forestry (non-fertilized poplar, N-fertilized poplar, non-fertilized Robinia, and poplar mixed intercropping with Robinia) as a randomized split-block design. Therefore, the objectives of the present study were (i) to obtain year-round N_2O emission data in various agro-ecosystems (cropland and agro-forestry) in central Germany and (ii) to provide more knowledge on how a legume-based crop production system may affect N_2O budget.

Methods

Field experiment

A field experiment was conducted at the research farm Reinshof (51.49° N, 9.93° E, 150 m asl) of the Georg-August-University Göttingen, Germany. The continental climate leads to an average precipitation of 651 mm and a mean temperature of 9.2 °C. The rainfall and temperature at field site during the investigation period can be found in Fig. 1. Here, annual mean temperature and cumulative rainfall were 10.1 °C and 677 mm in 2014, respectively. The soil was classified as Haplic Luvisol according to the FAO classification system. At 0–30 cm soil section, the soil contained 15 % clay, 73 % silt, and 12 % sand with a pH of 6.7, and 0.1 % total N and 1.0 % total organic carbon content [30].

The 1-year field experiment (part of a large field trial) was set up in April 2014 with the aim of comparing N_2O fluxes from the cropland (non-fertilized wheat (WT), N-fertilized wheat (NWT), non-fertilized faba bean (FB), wheat mixed intercropping with faba bean (WFB)), agro-forestry (non-fertilized poplar (PL), N-fertilized poplar (NPL), non-fertilized Robinia (RB), and poplar mixed intercropping with Robinia (PRB)) as a randomized split-block design. In the first year of the study, spring wheat (*Triticum aestivum* L.) cultivar Tybald and faba bean (*V. faba* L.) cultivar Fuego were sown at the same time at the optimum planting date for



spring wheat (see Table 1 for details). Agro-forestry treatments consisted of poplar (*Populus* “Hybride 275”) (PL); poplar mixed with Robinia (*R. pseudoacacia* “HKG 81901”) (PLR); Robinia (RB); and N-fertilized poplar (PLN). In the agro-forestry treatments, all trees were planted with 1 tree m⁻² on 30 April 2014. Plot size was 5 × 5 m for forest and 3 × 9 m for arable treatments with three replications for each treatment. After the harvest of crop land in 2014, winter wheat (*T. aestivum* “genus”), wheat intercropped with winter bean (*V. faba* “S_004”), mono-winter bean, and N-fertilized wheat were seeded on 28 October 2014. Seeding density in the mono-cropped winter wheat plots was 320 plants m⁻², intercropped plots were alternately seeded with wheat density of 160 plants m⁻² and a bean density of 20 plants m⁻², and mono-bean plots had a density of 40 plants m⁻². Granular N fertilizer in the form of calcium-ammonium-nitrate was applied at a rate of 80 kg N ha⁻¹ as a single dressing to the soil surface on 15 May 2014 and 25 March 2015 in the respective treatments.

Soil mineral N

For the analysis of soil mineral N, soil from 0–15 cm depth was sampled extracted with a 0.0125 M CaCl₂ solution (1:5 w/v) and shaken for 1 h. The extracts were then filtered with Whatman 602 filter paper and stored at -20 °C until analysis. The extracts were analyzed colorimetrically for the concentrations of NO₃⁻ and NH₄⁺ using the San++ continuous flow analyzer (Skalar Analytical B.V., Breda, The Netherlands).

Trace gas flux measurement

After the N application, gas samples were taken daily for a period of 1 week, followed by intervals of 2–3 days

Table 1 Sowing dates and fertilizer application rate of each treatment

Abbreviation	Crop	Fertilizer (kg CAN-N ha ⁻¹)	Stand	Date of sowing spring and winter crops
WT	Spring wheat/winter wheat	None	Mono	25 March 2014/28 October 2014
NWT	Spring wheat + N/winter wheat + N	80	Mono	25 March 2014/28 October 2014
WFB	Mixed intercropping (wheat and faba bean) ^a	None	Mixed intercrop	25 March 2014/28 October 2014
FB	Field beans ^a	None	Mono	25 March 2014/28 October 2014
PL	Poplar	None	Mono	30 April 2014
NPL	Poplar	80	Mono	30 April 2014
PRB	Mixed planting Poplar/Robinia	None	Mixed planting	30 April 2014
RB	Robinia	None	Mono	30 April 2014

^aInformation was given for both spring and winter crops

until the end of the vegetation period and once per week during the winter period using the closed chamber method described by Hutchinson and Mosier [31]. On each of 24 plots, basal rings made of polyvinyl chloride (PVC) (height 10 cm, diameter 60 cm) were pressed 5 cm into the soil. For the measurements, PVC chambers with an inner diameter of 60 cm and a height of 30 cm were put onto the basal rings, and tightened by a butyl rubber band all around the junction. The chambers were closed for 40 min within the period of 10.00–14.00 h at each sampling day. Gas samples were taken from the chamber atmosphere at 0–20–40 min after closing the chamber using pre-evacuated 12-ml glass vials (Labco, Crewe, UK). The top of the PVC chambers were covered with white polystyrene to reflect solar radiation and have temperature stability within the chamber.

Concentrations of N₂O were analyzed by a Bruker gas chromatography system (456-GC, Bruker, Billerica, USA) by deploying an electron capture detector (ECD) for N₂O. Operating conditions for the GC were as follows: injector temperature 95 °C, column temperature 85 °C, and detector temperature 320 °C. Samples were introduced using a Gilson auto-sampler (GX-281) (Gilson Inc., Middleton, WI, USA). Data processing was performed using the CompassCDS (vers. 3.0) software.

Statistics

Daily N₂O flux rates for dates between sampling dates were calculated using linear interpolation, and annual cumulative N₂O emissions were calculated as the sum of all daily flux rates for the vegetation period of each crop and the entire investigation periods during March 2014–August 2015. We used general linear model and Tukey's test for pairwise comparison of cumulative N₂O flux between the treatments within each year. Statistical analyses were done using SPSS version 13.0.

Results and discussion

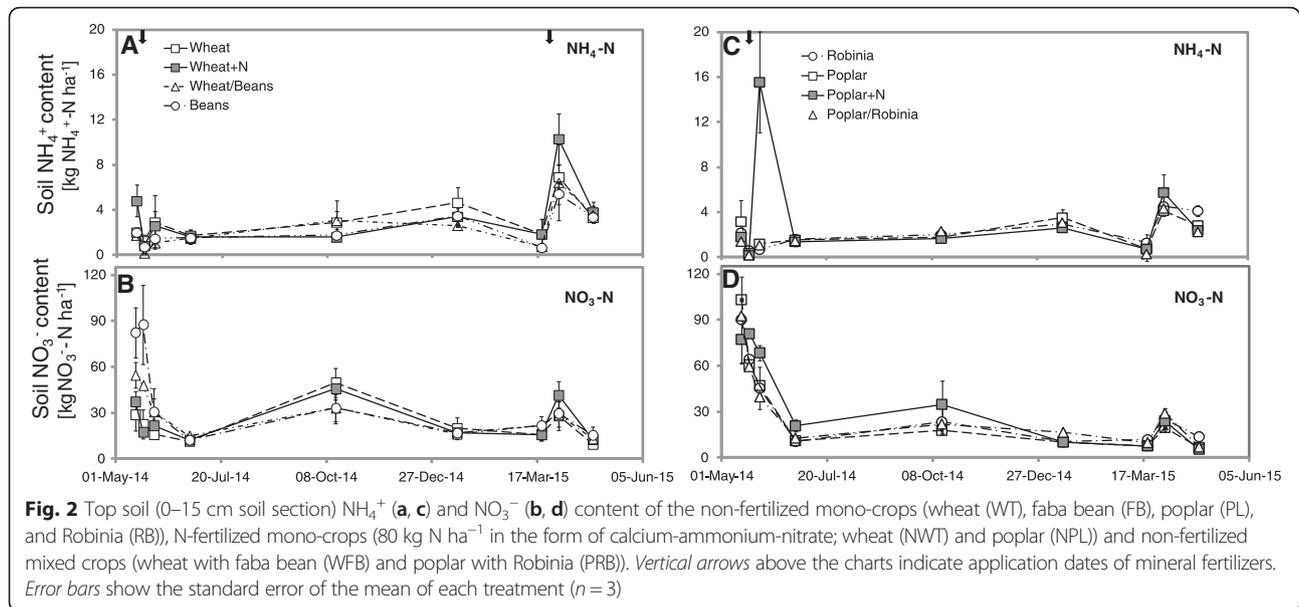
Rainfall and crop production

Rainfall at the site Reinshof was slightly over average during the period from 1 April to 1 July in 2014 (201 mm) and considerably below average during the period from 1 April to 1 July in 2015 (100 mm). Therefore, the 2014 spring period was much wetter than spring 2015. Overall, 2014–2015 winter period was quite mild with no significant freeze-thaw event.

Soil mineral N

The time course of the soil mineral-N concentrations are shown in Fig. 2. In soils planted with arable crops, soil NH₄⁺ concentrations during the investigation period remained rather low (below 10 kg N ha⁻¹ at 30 cm layer) in non-fertilized treatments. The application of mineral-N fertilizer in NWT treatment caused a slight increase in topsoil NH₄⁺ concentrations for a short time period which decreased rapidly to the background concentrations within a week. The low concentration of soil NH₄⁺ even in N-fertilized treatments (in the form of calcium-ammonium-nitrate) can be attributed to a rapid nitrification as soil texture and pH (6.7) serve ideal conditions for nitrification. In all treatments, there was a significant increase in topsoil NH₄⁺ concentrations in spring 2015 regardless from the N application. Early spring period in 2015 was reasonably dry compared to the same period in 2014. Therefore, the latter can be attributed to the processes related to the soil wetting after a long dry period (such as in spring 2015) which can accelerate N release from the mineralization of soil organic matter immediately after rewetting [32].

Overall treatments, soil NO₃⁻ concentrations in 0–15 cm soil segment varied between 10 and 85 kg NO₃⁻-N ha⁻¹. Here, NO₃⁻ was generally the dominant soil N form and highly variable when sampled soon after additions of fertilizer N. In all soils, concentrations of NO₃⁻ in the 0–15-cm layer decreased over time with the largest



decrease found in the arable crops specifically in WT and NWT treatments. During the vegetation period in 2014, soil NO_3^- content was generally higher in agro-forestry soils than in arable soils. Plant nutrient and water uptake was expected to be higher in cropland compared to the young agro-forestry treatments in 2014 due to small size and low growth rate of young trees. Thus, more rapid depletion of soil mineral N in arable crops than agro-forestry treatments can mainly be attributed to the differences in plant N uptake.

In arable land stand, soil NO_3^- concentrations were clearly higher (significant in 2014, $p < 0.01$) in FB than in other non-fertilized treatments during the vegetation period (Fig. 2a, b). For legume crops, inputs of biologically fixed N largely supplement to the uptake of soil mineral N to meet crop N demand. Thus, the legume species also take up mineral N from soils for growth before fixing additional N. The preferential use of soil mineral N helps explain why there is also significant depletion of soil NO_3^- in FB treatment [33]. In a review study, authors reported that average 41 % (for chickpea), 65 % (for faba bean), and 66 % (for field peas) of N that were present in legumes were derived from soil N [33]. However, slightly higher soil NO_3^- concentration in FB treatments than non-legume soils suggests that there were still reasonably more NO_3^- available for potential denitrification losses during the legume-growing season.

Seasonal N_2O emissions

In both years, flux data indicate that N_2O emissions were dominated by specific event periods (Fig. 3). Overall, maximum daily emissions of N_2O in the early summer

period in 2014 were 0.16 ± 0.07 and $0.04 \pm 0.01 \text{ kg N}_2\text{O-N ha}^{-1} \text{ day}^{-1}$ in agro-forestry and arable land treatments, respectively. In a 2-year field study, Lebender et al. [12] observed similar flux rates over a nearby site with similar soil conditions and agricultural practices (wheat and spring barley). Maximum N_2O emissions measured in agro-forestry treatments ($0.16 \pm 0.07 \text{ kg N}_2\text{O-N ha}^{-1} \text{ day}^{-1}$) have been usually observed in young agro-forest ecosystems [7, 27]. Almost all significant N_2O fluxes occurred as daily peak N_2O emissions and were measured only during the early summer period in 2014. The importance of these peak emissions in early summer period on the annual budget of N_2O emissions highlights the necessity of continuous flux monitoring to accurately determine the N loss from agro-ecosystems specifically in spring and early summer seasons [7, 20].

In 2014, N_2O emissions gradually decreased to the background levels (below $10 \text{ g N}_2\text{O-N ha}^{-1} \text{ day}^{-1}$) from the months of May to July and remained in background levels until March 2015 (Fig. 3). Interestingly, the latter was less than $0.01 \text{ kg N}_2\text{O-N ha}^{-1} \text{ day}^{-1}$ during the early summer period in 2015 for all treatments (including N-fertilized treatments). As seen in Fig. 1, early summer period in 2014 was relatively wet (April–June, 201 mm rainfall) compared to the same period in 2015 (April–June, only 100 mm rainfall). Therefore, we may attribute higher daily N_2O fluxes in 2014 than in 2015 (in early summer period) to the differences in mineral N and moisture content of the soil. Mineral N content of all soils in June 2014 was almost similar as compared to the same period in 2015, whereas N_2O fluxes were still about 10-fold higher in June 2014 than in June 2015. In this context, we may conclude that soil moisture seems to be the major

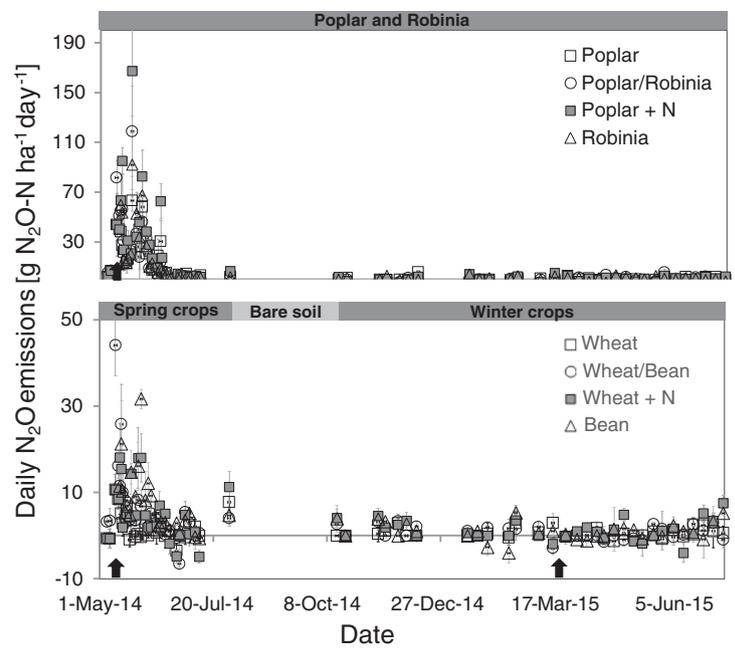


Fig. 3 Daily N_2O fluxes of the non-fertilized mono-crops (wheat (WT), faba bean (FB), poplar (PL), and Robinia (RB)), N-fertilized mono-crops (80 kg N ha^{-1} in the form of calcium-ammonium-nitrate; wheat (NWT) and poplar (NPL)), and non-fertilized mixed crops (wheat with faba bean (WFB) and poplar with Robinia (PRB)). Vertical arrows under the charts indicate application dates of mineral fertilizers. Horizontal bars above the charts indicate growing periods of crops. Error bars show the standard error of the mean of each treatment ($n = 3$)

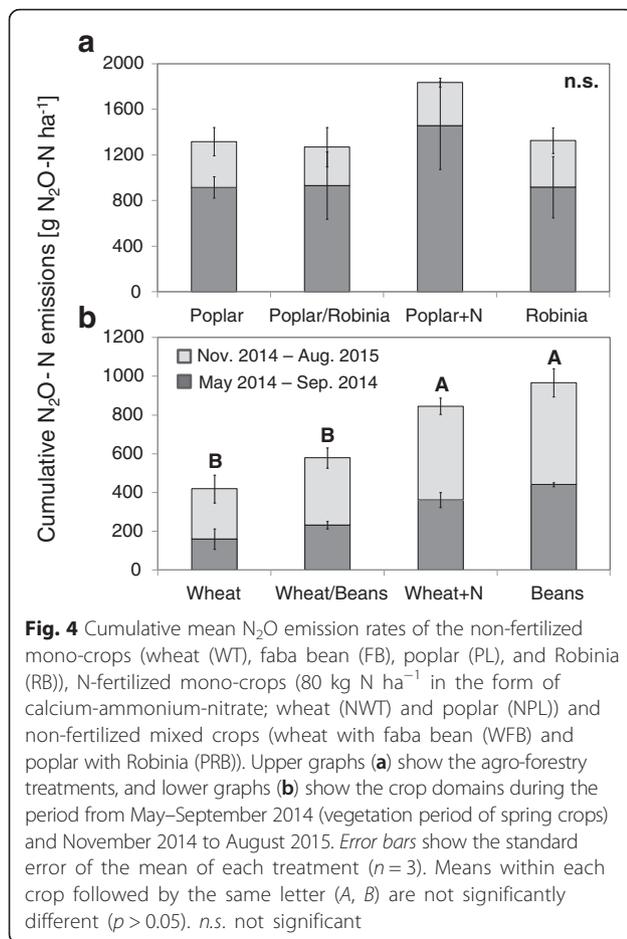
driving factor of higher N_2O emissions in the 2014 summer period than in 2015.

All the abovementioned factors (e.g., high moisture, high soil temperature in early summer, and moderate or high NO_3^- content of the soil) are known to trigger specifically the denitrification rate in soils. Thus, we may speculate that denitrification (fungal or bacterial) was the potential key source of measured large N_2O fluxes in the 2014 early summer period. Our earlier report supports this hypothesis in which sandy loam soil was incubated under laboratory conditions, and similar to the field experiment, large N_2O peak events were observed immediately after rewetting of the soil. Here, a stable isotope-labeling study clearly showed that denitrification was the major source (over 90 % of emitted N_2O) of large N_2O peaks that occurred in wet seasons [8, 19]. Furthermore, there is now a growing evidence that fungal denitrification may be the key process producing N_2O in such situations rather than bacterial denitrification [17]. Surely, more research is needed to reveal (i) the dominant processes and (ii) key microbial or fungal strains producing N_2O under these specific conditions (especially during early summer period).

Effect of plant species on N_2O emissions

Mean cumulative N_2O fluxes during the vegetation period in 2014 were 152 ± 58 , 217 ± 29 , and $441 \pm 10 \text{ g}$

$N_2O-N \text{ ha}^{-1}$ in WT, WFB, and FB treatments, respectively. Among the non-fertilized arable crops (wheat, wheat mixed intercropped with bean, and bean), N_2O emission over the 2014 growing seasons is highest in soils when faba bean (FB) was grown (Fig. 4). Introducing N-fixing legumes into cereal-based crop rotations may reduce synthetic mineral-N fertilizer use and thought to mitigate N_2O fluxes. However, the present study clearly showed that when faba bean was grown as a mono-crop, N_2O fluxes were about threefold higher compared to WT treatment. In contrast to the present study, authors reported that growing season N_2O emissions from N_2 -fixing legumes are significantly lower than from non-legumes and are often comparable to unfertilized background emissions [26, 34]. In line with the present study, Rochette and Janzen [26] (in a review study) concluded that legumes can produce substantial N_2O emissions. They speculated that the main source of N_2O emissions from soils planted with N_2 -fixing legumes during the vegetation period may be attributable to the N release from root exudates and/or from the decomposition of dead root residues. An alternative process that may contribute to the latter would be the N_2O emission during the N_2 -fixation process in the nodules where N_2 is fixed. Authors reported that several Rhizobium species in the free-living forms or in legume roots can denitrify NO_3^- and release N_2O from active



nodules most likely to prevent excess NO₃⁻ that inhibits the activity of N₂-fixing enzymes [35].

In agro-forestry treatments, both daily and cumulative N₂O emissions did not differ among each other in the 2014 growing period. Here, mean seasonal N₂O emissions (during the growing season of arable crops) were 1121 ± 161, 1102 ± 159, and 1052 ± 266 g N₂O–N ha⁻¹ in PL, PRB, and RB treatments (no significant difference), respectively. The cumulative mean N₂O fluxes during the 2014 growth period were considerably higher in agro-forestry than in arable crop treatments. During the first year, young plantations in agro-forestry domains generally have limited N and water uptake, while wheat and faba bean as arable crops are at their most productive growth stage specifically during May and June (growth rates are almost at their maximum during this period). Here, soil conditions seem to be more favorable specifically for denitrification in agro-forestry treatments than in soils planted with arable crops that may explain large N₂O emissions [27]. The cumulative mean N₂O emissions in agro-forestry treatments were about fivefold higher than in both WT and WFB treatments, and the

latter was still more than twofold higher when compared to FB plots. In line with the present data set, authors reported large N₂O fluxes after conversion of pastures lands [7] or grasslands [36]. Here, authors attributed large N₂O fluxes to the soil disturbance associated with tillage and cultivation that accelerate soil organic matter decomposition and microbial activity (nitrification and denitrification) leading to N₂O emissions. In the present experiment, soil tillage has been done almost at the same time for all treatments. Thus, we may speculate that the difference in N₂O emissions when comparing arable land to agro-forestry was mainly due to differences in water and nutrient uptake of plant species. Water and nutrient demand of young plantations during the first year are generally low which in return may cause more favorable conditions specifically for denitrification and N₂O losses from denitrification. Overall, we can therefore summarize that direct plant effect seems to be one of the key variables that regulates N₂O losses from soils. Zona et al. (2013) concluded that vegetation uptake of NO₃⁻ together with water ultimately may reduce the anaerobic volume of soils and may lower both denitrification rate and product stoichiometry of denitrification (lower N₂O/N₂O + N₂ ratio; meaning reduced N₂O and enhanced N₂ production) in agricultural soils [8, 17].

Effect of mineral N supply on N₂O emissions

Although it was not the main goal of the present experiment to study the effect of mineral-N addition on N₂O fluxes, we added fertilizer N to mono-crop wheat (80 kg N ha⁻¹; calcium-ammonium-nitrate) in parallel plots to be able to compare N₂O fluxes from soils planted with N₂-fixing plants (faba bean mono-culture or faba bean intercropped with wheat) with fertilized and non-fertilized wheat soils. N fertilization during the first year of new agro-forestry plantations is also not a common practice. However, N doses similar to the arable treatments were applied at the same date in order to be able to gain better scientific knowledge about the dominant factors regulating N₂O fluxes in agro-forest ecosystems. Expectedly, in all N-fertilizer treatments, N₂O fluxes increased immediately after fertilizer application, however, only in 2014 (wet early summer) but no response observed in 2015 (dry early summer). The latter clearly suggests that environmental factors specifically soil moisture was the most dominant factor in 2014 that leads to relatively high N₂O fluxes and without sufficient moisture or rainfall, fertilizer application alone does not affect N₂O fluxes significantly, e.g., in 2015. In line with the present study, authors also reported that the application of organic or inorganic fertilizers affects N₂O fluxes only in wet seasons but not in dry years [10, 17, 19].

Overall, the application of nitrogenous fertilizer affected N_2O fluxes predominantly in cropland soils and had limited impact in agro-forestry soils. Here, cumulative N_2O fluxes (from May to September 2014) were 46 and 121 % higher in fertilized than in non-fertilized agro-forestry (non-fertilized poplar vs. fertilized poplar) or in cropland treatments (non-fertilized wheat vs. fertilized wheat), respectively. The latter clearly suggests that N_2O fluxes were more dependent on soil mineral-N content in arable crops than in agro-forestry most likely due to greater competition between plants and N_2O -producing soil microorganisms in cropland than in agro-forestry soils. Fertilizer-derived cumulative N_2O emissions (emission factor) during the period from April to December 2014 were 0.21 and 0.45 % of applied N in wheat and poplar soils, respectively. Measured emission factors were significantly lower than what IPCC predicts (1 %; [4]). However, the latter was similar to what we reported in our previous study for central and northern Germany ([12, 19]). For operational reasons in the present IPCC protocol, the N_2O emission factor was set to 1 % for all fertilizer N regardless of crop or soil type. Large variations in N_2O emissions from different agricultural systems due to differences in management, climate, and soil type are very well known. Low N_2O emission factors in the present experiment and in our previous reports suggest that in the future, different emission factors should be considered at least for different crops or regions that account for their different risks of N_2O emissions.

Cumulative mean N_2O emission during the growing season was still 31 % higher ($p < 0.05$) in FB than in NWT treatment. In a review study, Rochette and Jansen [26] summarized that legumes can increase N_2O emissions during growth compared to evenly fertilized arable crops most likely due to the N release from the root exudates and decomposition of crop residues. Our study clearly agrees with Rochette and Jansen [26] and many others (e.g., [7, 27]) that growing legumes as mono-crop can increase N_2O fluxes compared to N-fertilized arable crops. On the other hand, seasonal N_2O fluxes were 35 % lower in WFB (wheat mixed intercropped with faba bean) than in NWT (wheat as mono-crop) treatment. The latter suggests that using legume crops as intercrop or mixed crop in wheat may significantly mitigate fertilizer-derived N_2O fluxes. However, surely more research is needed to upscale current findings due to the complexity and variability of N_2O fluxes in complex agricultural systems, e.g., mixed cropping systems.

Conclusions

The N_2O emission from soils is variable in space and time, thus measuring and quantifying variance in N_2O emissions is rather difficult, and there are only few field experiments available allowing long-term comparison of

various plant species (crops, legumes, and agro-forests) simultaneously with fertilizer effects. We see three take home messages:

- Currently, biogas production from energy crops is mainly based on anaerobic fermentation of mono-crops; however, high-yielding mono-crops require high N-fertilizer input that increases the risk of N_2O losses. Present study clearly showed that mixed intercropping agricultural systems (legume and non-legume plant species) may significantly lower (about 35 %) N_2O losses compared to the N-fertilized mono-crops.
- Cumulative N_2O emissions in agro-forestry soils were about 2–5-fold higher than in cropland soils. Soil conditions in agro-forestry treatments seem to be more favorable specifically for denitrification (due to limited water and nutrient uptake of young plantations during their initial growth stage) than soils planted with arable crops that may be responsible for large N_2O emissions.
- Cumulative mean N_2O emissions during the growth period of annual crops were 31 % higher ($p < 0.05$) in soils planted with faba bean than in N-fertilized wheat. We can conclude that legumes (when grown alone) can produce substantial N_2O emissions most likely due to enhanced denitrification activity in their rhizosphere due to N-rich root exudates/dead organic matter.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

MS contributed to the acquisition of funding, supervised the field work, perform the statistics, and drafted the paper (50 %). CW as a MSc student run the experiment (70 %; gas sampling, soil sampling, and analysis), collected and analyzed the data, and drafted the paper (10 %). AL as a PhD student contributed to the field experiment (crop management, soil sampling, and data analysis), and drafted the paper (5 %). JJ contributed to the acquisition of funding and design of the experiment and drafted the paper (5 %). HS contributed to the acquisition of funding, participated to the design of the experiment, and drafted the paper (5 %). CK drafted the paper and contributed to the planning of the experiment. SK as a PhD student contributed to the field experiment (crop management, soil sampling, and analysis), supervised the MSc student, and drafted the paper (25 %). All authors read and approved the final manuscript.

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